

IMPROVED LIGAND BINDING ASSAYS FOR VANILLOID RECEPTORS

BACKGROUND OF THE INVENTION

5 This application claims priority from U.S. Provisional Patent Application 60/240,628 filed October 16, 2000 and entitled "Improved Ligand Binding Assays for Vanilloid Receptors.

10 Noxious chemical, thermal and mechanical stimuli excite peripheral nerve endings of small diameter sensory neurons (nociceptors) in sensory ganglia (eg., dorsal root, nodose and trigeminal ganglia) and initiate signals that are perceived as pain. These neurons are crucial for the detection of harmful or potentially harmful stimuli (for example heat), tissue damage caused by local tissue acidosis, and physical movement (for example tissue stretch) that arise from changes in the extracellular space during inflammatory or ischaemic
15 conditions (Wall and Melzack, 1994).

20 Capsaicin (8-methyl-N-vanillyl-6-nonenamide), the main pungent ingredient in "hot" capsicum peppers, and its analogs interact at specific membrane recognition sites called vanilloid receptors. These receptors are expressed almost exclusively by primary sensory neurons involved in nociception and neurogenic inflammation (Bevan and Szolcsanyi, 1990). Capsaicin is a very selective activator of thinly or unmyelinated nociceptive afferents (Szolcsanyi, 1993; Szolcsanyi, 1996). Capsaicin can be blocked by a selective antagonist, capsazepine. Another ligand is the potent tricyclic diterpene resiniferatoxin (RTX), (Szolcsanyi et al., 1991), a molecule that binds
25 with nanomolar affinity at the capsaicin-binding site.

30 Recently, one receptor for capsaicin (VR1) was cloned from rat (Caterina et al., 1997) and shown to be a coincidence detector for H⁺ (low pH) and heat (Tominaga et al., 1998). VR1 is expressed in small nociceptive neurons of the dorsal root ganglion, consistent with its role in modulating peripheral pain (Tominaga et al., 1998). VR1 is a ligand-gated non-selective cation channel that shows pronounced outward rectification (Caterina et al., 1997). The vanilloid ("capsaicin") receptor VR1 is activated by capsaicin and

RTX, and activation of VR1 is blocked by the antagonists capsazepine (CPZ); (Bevan et al., 1992) and ruthenium red (RR; (Wood et al., 1988)). Recently, rat VR1 and VR2 and a partial cDNA sequence of human sequences were disclosed in the WIPO publication WO 99/09140.

5 The densities of VR1 receptors can be tested using a [³H]RTX binding assay (Szallasi and Blumberg, 1990; Szallasi and Blumberg, 1993). Indeed, high expression of VR1 receptors was observed in rat and human spinal cord and dorsal root ganglia (Szallasi et al, 1993; Szallasi and Goso, 1994; Acs et al., 1994). Protons inhibited [³H]RTX binding to VR1 receptors (Szallasi et al.
10 1995).

 Prior ligand binding assays using the VR-1 receptor teach that the pH must be near physiological conditions. In these assays, ligand binding was reduced by 50% and 70% at pH 8.0 and pH 9.0, respectively (Szallasi and Blumberg, 1993).

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SUMMARY OF THE INVENTION

 In contrast to what is suggested in the art, the present invention provides the surprising discovery that the binding capacity of certain ligands of the
20 Vanilloid receptor increases at pH values that are greater than pH 7.4. The present invention provides improved assays to measure competitive vanilloid receptor binding of a known radiolabeled ligand and a test compound binding in aqueous buffers at a pH in the range of about 7.5 – 10.0. The present invention also provides the discovery that divalent cations also increase the
25 binding capacity of certain ligands for the Vanilloid receptor. Therefore the aqueous solutions used for the methods of the present invention advantageously may include, as one component, a divalent cation.

 The methods of the present invention are useful to find compounds that bind to Vanilloid receptors.

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BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1. Effect of pH on [³H]RTX binding to the hVR1 receptor. Cell membranes (60 µg protein/ml) were incubated with [³H]RTX (0.4 nM) in buffer samples with differing pH at 25 C for 60 min. The results are representative of two experiments with each point assayed in triplicate.

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Figure 2. pH changes affinity of [³H]RTX for the hVR1 receptor.

Figure2A Cell membranes (60 µg protein/ml) were incubated with varying concentrations of [³H]RTX in buffer samples with differing pH at 25°C for 60 min. (pH 5.2, pH 7.4 and pH 8.6)

10 Figure 2B: Cell membranes (60 µg protein/ml) were incubated with varying concentrations of [³H]RTX in buffer samples with differing pH at 25°C for 60 min. (pH 7.4, pH 8.6 and pH 9.6).

Figure 3. Effect of vanilloid analogs on [³H]RTX binding to the hVR1 receptor

15 **at pH 7.4 and pH 8.6.** Membranes were incubated with [³H]RTX (0.4 nM) and varying concentrations of vanilloid analogs at 37°C for 60 min. The data are representative of two experiments with each point assayed in duplicate. The results demonstrate that vanilloid analogs used in this study dose-dependently inhibited [³H]RTX binding at both pH 7.4 (Fig. 3A) and pH 8.6 (Fig. 3B). The

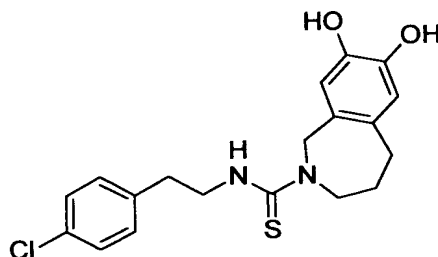
20 EC₅₀ values of RTX and capsaicin were slightly decreased from pH 7.4 to pH 8.6. In contrast, the EC₅₀ value of capsazepine was significantly increased.

Figure 4. Calcium and magnesium increased [³H]RTX binding. Membranes were incubated with [³H]RTX (0.25 nM) at pH 8. Without calcium and

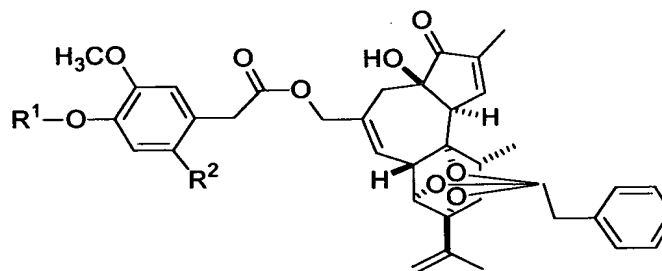
25 magnesium the signal was decreased by 20%. EGTA (10 mM) inhibited [³H]RTX binding by 70%.

DETAILED DESCRIPTION OF THE INVENTION

30 Capsaicin is a compound of the formula:



Resiniferatoxin is a compound of the formula:



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- (d) determining if the test compound bound to the receptor by observing a reduction in the amount of expected labeled ligand.

The methods can optionally include a step of removing unbound labeled ligand from the solution and also optionally the step of isolating the receptor protein.

The aqueous solution of the present invention may be composed of any buffering species that provides a suitable pH. The choice of a buffer to provide suitable pH is well known in the art. The pH suitable for the methods of the present invention are in the range of about 7.5 to about 10.0, preferably from about pH 8.0 to about 9.5, more preferably from about pH 8.1 to about 9.1, and particularly at about pH 8.6.

There are a variety of buffers well known in the art that can be used for the methods of this invention. A preferred buffer is HEPES (N-[2-hydroxyethyl]piperazine-N'-2-ethanesulfonic acid with a pka of about 7.55. Other buffers include, but are not limited to MES (morpholinoethane sulfonic acid, pka about 6.2); MOPS (morpholinopropane sulfonic acid, pka about 7.2); PIPES (Piperazine-N,N'-bis(2-ethane sulfonic acid, pka about 6.8); and TES (N-Tris(hydroxymethyl)methyl-2-aminoethane sulfonic acid; pka about 7.5).

The solution may contain agents that minimize protein adsorption onto the surface of the vessel containing the solution. Such agents are well known and include for example, a protein such as bovine serum albumin or immunoglobulin, or an amino acid, such as glycine.

Advantageously, the solution may contain a divalent cation. Use of a divalent cation has been demonstrated here to enhance ligand interaction with the vanilloid receptor. Particularly preferred divalent cations are Magnesium and Calcium. Other divalent cations can be tested and used in these assays without undue experimentation. Divalent cations are preferably used at a concentration in the range of about 0.1mM to about 10mM. Agents that chelate divalent cations, such as EDTA or EGTA, are preferably not used in the aqueous solution.

The term "test compound" is used herein to refer to a candidate molecule having the potential capacity to interfere with the binding of a labeled ligand and the ligand-interacting portion of a vanilloid receptor.

5 The term "labeled ligand" as used herein in connection with the assays of this invention is a ligand known to bind to the vanilloid receptor protein, which has a detectable label including, but not limited to, a fluorescent molecule or a radioactive tag. Examples of fluorescent molecules suitable for use in the present invention include, but are not limited to, coumarins, xanthene dyes such as fluoresceines, rhodols, and rhodamines, resorufins, cyanine dyes
10 bimanes, acridines, isoindols, dansyl dyes, aminophthalic hydrazides such as luminol and isoluminol derivatives, aminophthalimides, aminonaphthalimides, aminobenzofurans, aminoquinolines, dicanohydroquinones, and europium and terbium complexes and related compounds. The types of radioactive tags used to label the ligand include any of a variety of known β -particle emitters or Auger
15 electrons, including [^3H], [^{14}C], [^{35}S], [^{33}P], [^{32}P], [^{125}I], and [^{131}I], with [^3H] being generally preferred due to its relative safety. In a most preferred embodiment of the present invention, the concentration of labeled ligand used is closely matched to the natural ligand's affinity (K_d) for its receptor. A preferred labeled ligand is resiniferatoxin or RTX, of which tritiated forms are well known.

20 The term "ligand-interacting portion of a vanilloid receptor protein" refers to that region(s) of a vanilloid receptor protein that interacts with the ligand being used in the assay. Proteins are typically divided into functional regions including transmembrane regions, one or more binding domains, intracellular regions, extracellular regions, regions that include particular folding
25 characteristics and the like. Those of ordinary skill in the art are able to create truncated fragments, receptor protein with altered sequences and chimeric proteins that can be used to define these functional regions. In this case it is contemplated that the assay incorporate at least that portion of the vanilloid receptor that binds to the ligand used in the assay.

30 Vanilloid receptors suitable for the methods of the present invention include receptors derived from any mammal, particularly human, mouse, rat, and monkey. There are several distinct genes that encode different vanilloid

receptor proteins. A number of those are referenced in the publications cited herein. The preferred vanilloid receptors include those that bind resiniferatoxin including, but not limited to, VR-1. The VR-1 receptor may be obtained using methods well known in the art including using the human VR-1 sequence (as provided as GenBank accession number NM_018727).

The vanilloid receptors can be obtained from a number of sources. In one example, the vanilloid receptors are isolated from native cells, for example, but not limited to, dorsal root ganglia expressing the vanilloid receptors such as described by Szallasi and Blumberg, 1993. In another embodiment the vanilloid receptors are obtained from cells expressing a cDNA encoding a recombinant vanilloid receptor. Preferably, at least the ligand-interacting portion of the vanilloid receptor protein is used. However, the entire protein may be used or the ligand-interacting portion of the receptor protein may be combined with other portions of other proteins, for example, one or more membrane-binding domains from other proteins. These chimeric protein still retain vanilloid receptor protein ligand-binding characteristics.

Following the formation of the aqueous solution of step (a), the solution is incubated for a time sufficient to allow the ligand and the vanilloid receptor or the test compound and the vanilloid receptor to come into contact. Methods for determining a suitable incubation time can be determined using the examples as described herein.

Next, in a preferred embodiment, unbound labeled ligand is removed from the solution. Methods for removing unbound labeled ligand from the solution can be performed using any of a variety of techniques known in the art, such as suitable adsorption strategies, membrane separation techniques where the vanilloid receptor protein is membrane bound or through the use of molecules such as alpha 1 acid glycoprotein, and the like.

In a further step of the assay of this invention, the receptor protein is isolated from the aqueous solution. In one embodiment, the ligand binding domain of the vanilloid receptor protein is associated with a membrane, such as cellular membrane or artificial membrane preparations. In another embodiment, the vanilloid receptor is created as a soluble protein. Methods for

removing membrane or isolating receptor protein are known in the art and include, for example, selective centrifugation methods, adsorption steps, column chromatography, antibody-mediated precipitation, and the like.

Preferably the methods of the assay of this invention are performed in order, however, those of ordinary skill in the art will understand that, as one example, the removing step and the isolating step may be combined as one step or performed in any suitable order that facilitates removal of unbound label from labeled receptor protein. Thus, in one assay the removing step may be performed before the isolating step, while in another assay, the format of the assay may be better performed if the isolating step and removing step are combined as a single step.

As a final step in the assay of this invention, suitable calculations and comparisons are made, using the appropriate controls, and the like to determine whether or not the test compound has bound to the ligand-interacting portion of the vanilloid receptor. In a preferred example, suitable controls are included in the assay that do not include test compound and permit a comparison between controls that do not include test compound and samples including test compound. A reduction in the amount of expected labeled ligand is indicative of test compound binding.

In a preferred assay of this invention, the ligand-interacting portion of the vanilloid receptor protein is associated with cell membrane and the isolating the receptor protein step comprises removing membrane from the aqueous solution.

The invention can be better understood by way of the following examples. These examples are representative of the preferred embodiments, but are not to be construed as limiting the scope of the invention.

EXAMPLE1

MATERIALS

Resiniferatoxin, capsaicin and capsazepine were purchased from Research Biochemical International (Natick, MA). HEPES and CASPO were purchased

from Sigma (St. Louis, MO). [^3H]resiniferatoxin (RTX) was purchased from NEN (Boston, MA). HEK 293 cells were transfected with the human vanilloid receptor(VR1).

5 METHODS

Cell culture. HEK 293 cells were grown as monolayers in DMEM (GIBCO) containing 10% fetal bovine serum and 1xPSA (Cascade Biologics) in an incubator with an atmosphere of 5% CO_2 at 37°C. HEK 293-hVR1 cells were grown in the same media containing 200 $\mu\text{g/ml}$ of zeocin (Invitrogen).

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Membrane preparation. Cells were washed with Hank's Balanced Salt Solution, dissociated with cell dissociation buffer (Sigma), and then centrifuged at 1000 x g for 5 min. Cell pellets were homogenized in cold 20 mM HEPES buffer, pH 7.4, containing 5.8 mM NaCl, 320 mM sucrose, 2 mM MgCl_2 , 0.75 CaCl_2 and 5

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mM KCl and centrifuged at 1000 x g for 15 min. The resultant supernate was then centrifuged at 4000 x g for 15 min. The pelleted membranes were kept in a -80°C freezer.

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[^3H]RTX binding assay. The assay procedure was modified from that described previously (Szallasi and Blumberg, 1993). About 120 μg protein/ml from membranes were incubated with indicated concentrations of [^3H]RTX in 0.5 ml of the HEPES buffer (pH 4.1 to pH 8.6) or CASPO buffer (pH 8.6 to pH 10.6) containing 0.25 mg/ml fatty acid-free bovine serum albumin at 37°C for 60 min. The reaction mixture was then cooled to 4°C, and 0.1 mg α_1 -acid glycoprotein

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was added to each sample and incubated at 4°C for 15 min. The samples were centrifuged at 18500 x g for 15 min. The tip of the microcentrifuge tube containing the pellet was cut off. Non-specific binding was tested in the presence of 200 nM unlabeled RTX. Bound radioactivity was quantified by scintillation counting.

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RESULTS

Effect of pH on [^3H]RTX binding to hVR1 receptors. Protons are known to stimulate calcium influx via the VR1 receptor. To study whether protons affect [^3H]RTX binding, membranes were incubated with [^3H]RTX at various pH values from 4.2 to 10.6. The results showed a biphasic effect (Fig. 1). [^3H]RTX binding increased from pH 4.2 to pH 8.6, but decreased from pH 8.6 to pH 10.6. The non-specific binding did not change significantly.

Mechanisms of pH affecting [^3H]RTX binding. To investigate whether the pH changes observed resulted from changes in the binding affinity or the apparent density of the binding sites, we performed saturation binding of [^3H]RTX at pH 5.2, pH 7.4, pH 8.6 and pH 9.6 (Fig. 2). The data are representative of two experiments with each point assayed in duplicate. The results demonstrated that the affinity (K_d values) of [^3H]RTX for hVR1 receptors was increased with increasing pH from 5.2 to 8.6 without a change in the number of binding sites (B_{\max}), whereas the affinity was decreased with increasing pH from 8.6 to 9.6 with a decrease in number of binding sites.

The K_d values of [^3H]RTX and the B_{\max} values are summarized in Table 1.

Table 1. K_d and B_{\max} values of [^3H]RTX binding to hVR1 receptor in buffers with different pH value.

pH	K_d (nM)	B_{\max} (fmol/mg protein)
pH 5.1	6.62 ± 5.58	ND
pH 7.4	0.65 ± 0.12	928 ± 53
pH 8.6	0.18 ± 0.04	869 ± 41

pH9.6

 0.60 ± 0.31 661 ± 127

K_d and B_{max} values were obtained from Figure 2. ND: not determinable.

Effect of pH on vanilloid ligand binding to hVR1 receptors. A number of vanilloid

5 ligands were tested for their ability to inhibit the binding of [3 H]RTX to hVR1 receptors in pH 7.4 and pH 8.6 buffer. In pH 7.4 buffer, competition for [3 H]RTX was in the order: RTX>> capsaicin = capsazepine (Fig. 3a). Similarly, in pH 8.6 buffer, competition for [3 H]RTX was in the order: RTX>> capsaicin > capsazepine (Fig. 3b). The IC_{50} values of RTX and capsaicin were slightly
10 decreased from pH 7.4 to pH 8.6 (Table 2). The IC_{50} value of capsazepine was significantly increased from pH 7.4 to pH 8.6 (Table 2). A yellow color is seen in membrane pellets from capsazepine at pH 8.6, suggesting that capsazepine might be oxidized from its double phenol structure to a double quinol structure.

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Table 2. IC_{50} values of vanilloid analogs which inhibit [3 H]RTX binding to hVR1 receptor in buffers with different pH value.

Buffer PH	IC_{50} (nM)		
	Resiniferatoxin	Capsaicin	Capsazepine
PH7.4	0.78 ± 0.15	630 ± 202	206 ± 43
PH8.6	0.25 ± 0.03	256 ± 37	$>10,000$

IC₅₀ values were obtained from Figure 3.

5 Effect of calcium and magnesium on [³H]RTX binding. The addition of calcium
and magnesium to the assay were found to increase binding and were used to
further optimize the assay. The binding assay was performed as described
earlier using pH8.0 buffer with the inclusion or chelation of divalent cations. As
seen in Figure 4, the presence of either 0.75mM CaCl₂ or 2mM MgCl₂
increased the total ligand binding compared with the buffer lacking the divalent
10 cations without increasing the nonspecific ligand binding. The presence of both
cations increased the ligand binding. In contrast, the presence of a divalent
cation chelator, EGTA, reduced the total amount of ligand binding.

15 There will be various modifications, improvements, and applications of
the disclosed invention that will be apparent to those skilled in the art, and the
present disclosure is intended to cover such embodiments. Although the
present invention has been described in the context of certain preferred
embodiments, it is intended that the full scope of the disclosure be measured
20 by reference to the following claims.

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